

Development of compact transcriptional effectors using high-throughput measurements in diverse contexts

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Transcriptional effectors are protein domains known to activate or repress gene expression; however, a systematic understanding of which effector domains regulate transcription across genomic, cell type and DNA-binding domain (DBD) contexts is lacking. Here we develop dCas9-mediated high-throughput recruitment (HT-recruit), a pooled screening method for quantifying effector function at endogenous target genes and test effector function for a library containing 5,092 nuclear protein Pfam domains across varied contexts. We also map context dependencies of effectors drawn from unannotated protein regions using a larger library tiling chromatin regulators and transcription factors. We find that many effectors depend on target and DBD contexts, such as HLH domains that can act as either activators or repressors. To enable efficient perturbations, we select context-robust domains, including ZNF705 KRAB, that improve CRISPRi tools to silence promoters and enhancers. We engineer a compact human activator called NFZ, by combining NCOA3, FOXO3 and ZNF473 domains, which enables efficient CRISPRa with better viral delivery and inducible control of chimeric antigen receptor T cells.

Over 10% of human genes encode proteins that localize to the nucleus and function to regulate gene expression at the level of mRNA transcription^{1–3}. These transcription factors (TFs) and chromatin regulator proteins represent a reservoir of potentially useful effector domains for constructing transcriptional and epigenomic perturbation tools for synthetic biology and dissecting biological processes^{4–8}. Transcriptional effector domains, also known as activators and repressors, are the regions of these proteins that can increase or silence transcription of a gene upon recruitment to its promoter^{1,9}. When fused to DNA-binding domains (DBD), they can be used as tools to manipulate

the expression of endogenous genes and long noncoding RNAs^{5,10,11} and control synthetic gene circuits¹² or endogenous regulatory elements^{13,14}, which has therapeutic potential^{15–17}.

However, transcriptional effectors can function differently depending on the biological context of their recruitment to a gene. For example, targeting of effector domains to the same genomic context but in different cell lines^{18–20} or in different mouse developmental stages *in vivo*²¹ can result in variable transcriptional and epigenetic effects. The target locus of effector recruitment can also have a role^{22–25}, wherein recruitment of effector domains to different promoters and enhancers

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in the same cell type results in distinct effects on transcription^{13,18,19,26–28}. Finally, the DBD can affect effector function^{19,29}.

We currently lack a system-level understanding of how frequently effector function depends on the target, cell type or DBD context, which is critical for identifying effectors that are robust across these conditions. Recently, pooled screening methods that measure effector strength for large libraries of domains have greatly expanded the list of sequences known to function as effectors^{30–34}, but these scalable methods use synthetic reporter genes instead of targeting endogenous loci and have not yet been used to compare activities across many contexts. Meanwhile, other salient features like deliverability, size, expression level and off-target toxicities remain uncharacterized for the majority of effectors.

To address these questions, we performed high-throughput recruitment (HT-recruit) to measure the effector function of 5,092 Pfam-annotated domains from human nuclear proteins (the Pfam library) across 17 biological contexts and integrated this data with two pre-existing datasets³². By adapting HT-recruit to use the programmable DBD dCas9, we were able to target endogenous promoters and enhancers. Using these large-scale datasets, we identify forms of context-specificity, which varied by effector family and were striking in the helix-loop-helix (HLH) family of domains. We then confirmed and expanded these results with screens of 114,288 tiling sequences from all human chromatin regulators and TFs (the CRTF tiling library) across two endogenous gene contexts, which we integrated with three pre-existing reporter gene datasets³⁴. This comprehensive testing of short human effector domains in human cells across multiple contexts allowed us to select robust activators and repressors to be used in synthetic biology tools. Finally, with additional measurements of effector expression level, deliverability and cellular toxicity, we developed improved effectors for CRISPR interference (CRISPRi), CRISPR activation (CRISPRa) and synthetic TF circuits. For repression, we find Krüppel-associated box (KRAB) domains are the most potent, and from the entire set of 336 human KRAB domains, we find several top KRAB domains that outperform the traditionally used ZNF10 KRAB. From the activation domains, we constructed a short potent combination comprising NCOA3, FOXO3 and ZNF473 domains (NFZ) whose improved utility for CRISPRa and synthetic TFs we demonstrate in macrophages, induced pluripotent stem cells (iPSCs) and T cells.

Results

HT-recruit quantifies effector function across contexts

We set out to measure transcriptional effectors quantitatively across biological contexts by performing HT-recruit screens using two DBDs (rTetR and dCas9) to recruit effectors to various gene targets in two cell types (Fig. 1a and Extended Data Fig. 1a). First, to study different

promoter contexts while keeping other parameters constant, we developed a set of reporters with varied promoter origins and strengths (Fig. 1b). To study how effector–promoter interactions differ across cell types, we installed these reporters at the AAVS1 safe harbor in both K562 and HEK293T cells (Fig. 1a). As expected, all minimal promoters (minimal cytomegalovirus (minCMV), non-TATA box-containing X chromosome and chromosome 21 (NTX and NT21) promoters) started OFF and could be activated with the VP64 activator domain (Fig. 1b). Two of the constitutive promoters, pEF1α and UbC, started ON and could be repressed by the ZNF10 KRAB domain (Fig. 1b). The Rous sarcoma virus (RSV) promoter was rapidly silenced upon installation in both cell types (Extended Data Fig. 1b), so we did not use it for screens. In certain cases we observed promoter silencing to be cell-type specific—the phosphoglycerate kinase 1 (PGK) promoter was constitutively ON in K562 cells but was background silenced in HEK293T cells (Fig. 1b). Based on this observation, we decided to use PGK as a repressible promoter in K562 and as an activatable promoter in HEK293T cells (Fig. 1b). Overall, the expression levels for each promoter were similar across the two cell types ($R^2 = 0.86$) except for the PGK promoter (Extended Data Fig. 1c).

To extend HT-recruit to endogenous loci, we used dCas9 and targeted the promoters of endogenous genes encoding cell surface proteins. Targeting surface proteins allows us to use fluorescence antibodies to immunostain cells, thus providing a way to monitor single-cell gene expression variability during individual recruitment assays by flow cytometry and to magnetically separate a large number of ON and OFF cells during HT-recruit (Fig. 1a). For a repressible context, we targeted the highly expressed surface marker CD43 in K562 cells. First, we individually recruited either dCas9 alone or dCas9–KRAB with single guide RNAs (sgRNAs) targeting the CD43 transcriptional start site (TSS) and found two sgRNAs, sg10 and sg15, for which repression depended on the KRAB repressor. Similarly, we identified sgRNAs with which dCas9–VP64 or VPR could activate the lowly expressed *CD2*, *CD20* and *CD28* genes (Fig. 1c and Extended Data Fig. 1d,e).

Then, we generated lentiviral libraries with 5,092 Pfam domains from nuclear-localized human proteins and 499 random and 362 negative control sequences tiling the nonnuclear protein DMD (hereafter Pfam library) fused to rTetR or dCas9 and delivered them to cell lines with a low multiplicity of infection (MOI) such that most cells express one DBD-domain fusion. Using these components, we performed eight rTetR-targeted screens across six synthetic reporter contexts in two cell types and 11 dCas9-targeted screens with sgRNAs targeting endogenous genes plus two dCas9 screens at reporters using a sgRNA targeting the TetO motif. We also integrated data from two previous screens using rTetR and the Pfam library (Fig. 1d and Extended Data Fig. 1f,g). To measure the activities of sequences beyond annotated Pfam domains, we also used our recently designed 20× larger library

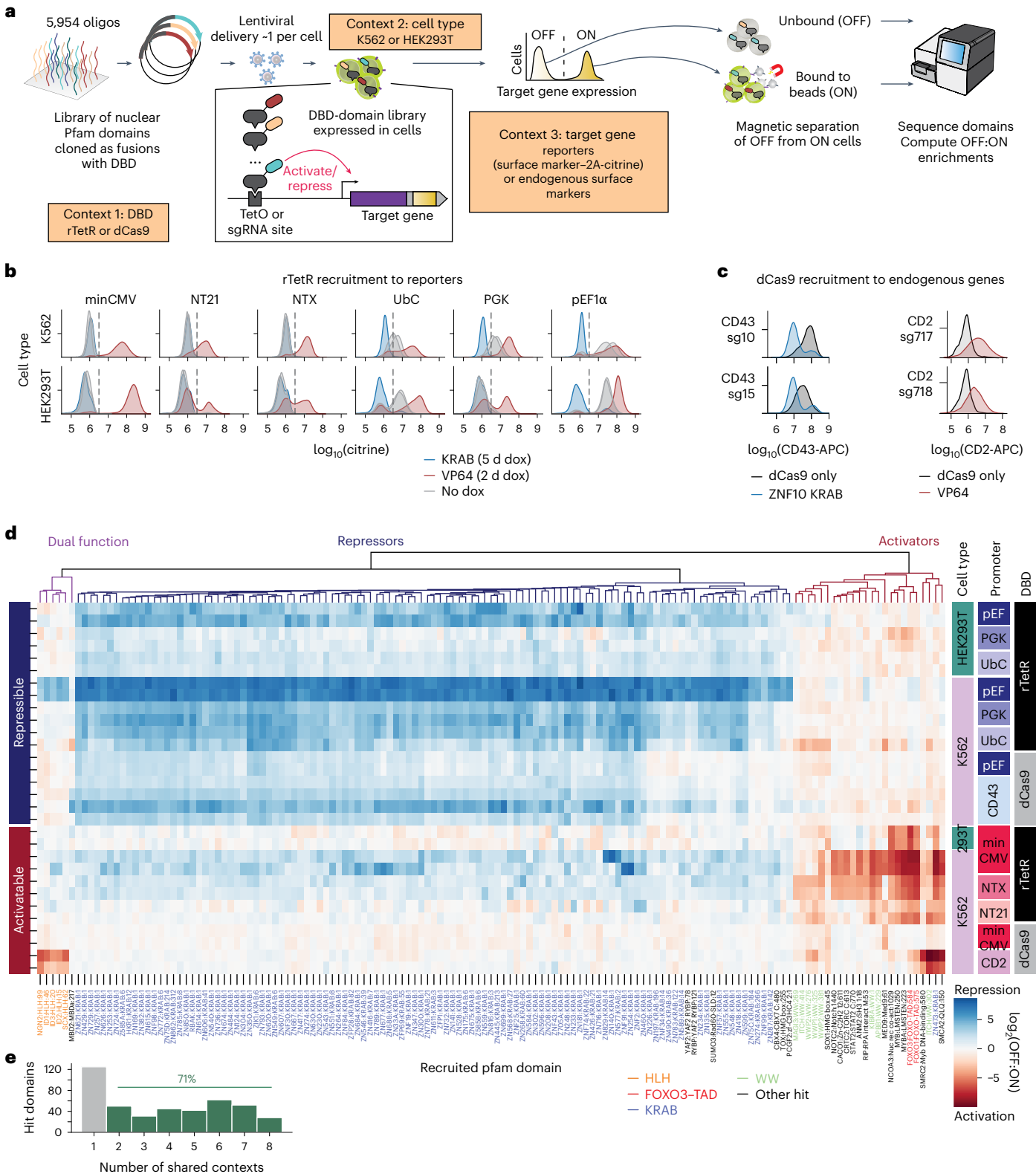
Fig. 1 | HT-recruit quantifies transcriptional effector functions across DBD, cell type and target gene contexts. a, Schematic representation of HT-recruit to quantify transcriptional effector function while varying the context of DBDs, cell type and target synthetic reporters or endogenous genes. A pooled library of Pfam domains from human nuclear proteins of ≤80 amino acids is synthesized as 300-mer DNA oligonucleotides, cloned downstream of the DBD (context 1) and delivered to cells (context 2) at a low MOI such that most cells express a single domain. The synthetic reporters encode a synthetic surface marker (lgx-hlg1-Fc-PDGRF) and fluorescent marker (citrine), while the endogenous target genes encode surface markers (context 3). After the recruitment of Pfam domains, ON and OFF cells were magnetically separated using beads that bind these synthetic or endogenous surface markers, and the domains were sequenced in the bound and unbound populations to compute enrichments. **b**, Expression of synthetic reporters. Positive control effectors, such as ZNF10 KRAB repressor or VP64 activator, were stably delivered by lentivirus. Cells were treated with 1,000 ng ml^{−1} doxycycline for 5 days for repression and 2 days for activation (or untreated as a negative control), and citrine expression was measured by flow cytometry after gating for rTetR delivery (mCherry⁺; $n = 2$ infection replicates

shown as histogram curves). **c**, Expression of endogenous surface marker genes in K562 cells measured by immunostaining and flow cytometry. dCas9 fusions and sgRNAs were delivered by lentivirus and selected for by blasticidin and puromycin, respectively. Data are gated for sgRNA delivery (mCherry⁺ in CD43 and GFP⁺ in CD2 samples) and for dCas9 (BFP⁺; $n = 1$ infection replicate). **d**, Transcriptional effector hits' activation and silencing activity across different target gene, DBD and cell-type recruitment contexts ($n = 2$ biological replicates per screen). A subset of effectors (columns) are shown that are a hit at a high threshold of 3 s.d. stronger than the median of the poorly expressed domains in ≥2 samples (rows; $n = 143$). Unbiased column clustering shows three major clusters of effectors (top). dCas9 targets pEF1α and minCMV with sgTetO-1, CD2 with sg717 and CD43 with sg10 (upper two rows) and sg15. Column labels on the bottom show the protein, Pfam domain and domain start position within the protein; select Pfam domain families are colored. Rows are manually ordered, with the targets that are predominantly repressible above the activatable targets. **e**, Distribution of the number of screen contexts in which a Pfam domain was a hit effector in two replicates. Domains that are shared hits in multiple contexts are colored green.

that tiles all human CRTFs in 80 amino acid tiles at ten amino acid steps ($n = 128,565$ elements, hereafter CRTF tiling library)³⁴ and recruited it with dCas9 to CD2 and CD43 in K562 cells (Extended Data Fig. 2a,b). We additionally integrated data from three previously performed rTetR screens with the CRTF library at pEF, PGK and minCMV³⁴.

To minimize confounding effects due to protein stability, we identified domains that were well-expressed in both cell types and when fused to either DBD. Expression was measured by permeabilizing

the cells, staining with an anti-FLAG antibody, sorting into high and low protein expression level bins and then sequencing the domains in those two cell populations ($R^2 = 0.44\text{--}0.78$ across comparisons of replicates; Extended Data Fig. 3a). Overall, domain expression was similarly correlated between cell types ($R^2 = 0.6$; Extended Data Fig. 3b) and whether they were fused to rTetR or dCas9 ($R^2 = 0.56$; Extended Data Fig. 3c,d). We labeled a domain as well-expressed if its score was 1σ above the median of the random controls. Meanwhile, the poorly



expressed domains, which included most random sequence controls, served as a large set of negative controls for activation and repression screens to set our hit threshold.

After filtering out poorly expressed domains, we conservatively called activation and repression hit domains with $\log_2(\text{OFF:ON})$ scores 2 s.d. beyond the median of poorly expressed elements (higher scores for repressors and lower for activators). This pipeline called 435 Pfam domains as hits in both replicates of a context, with 71% of these hits being rediscovered in multiple contexts (Fig. 1e and Supplementary Table 1). We also called 1,261 tiled regions as hit domains from the larger CRTF tiling library (Methods; Supplementary Tables 2 and 3). The hits clustered into the following three categories: activators, repressors and dual-function effectors that are context-dependent activators or repressors (Fig. 1d). We then set out to investigate the diverse categories of context-dependent and context-robust effectors.

Activator function across targets and cell types

We first considered activator promoter-specificity when using rTetR to target the Pfam library at minimal promoter reporters in K562 cells. We found that these core promoters responded very similarly to activator recruitment, wherein 85% of well-expressed NTX activators also activate minCMV, and do so with well-correlated strengths ($R^2 = 0.8$; Fig. 2a,b and Extended Data Fig. 4a,b), which we individually validated with flow cytometry. In contrast, only 9% of the well-expressed minCMV activators, such as FOXO3-TAD, were able to re-activate the background-silenced PGK promoter in HEK293T cells (Extended Data Fig. 4c,d).

To extend these experiments to endogenous gene targets, we used dCas9 recruitment. First, we targeted the minCMV reporter using a TetO-targeting sgRNA. dCas9 recruitment resulted in fewer hits than rTetR at the same reporter, perhaps due to weaker and/or lower copy number recruitment; however, the strongest hit, the Med9 mediator component, was validated individually (Extended Data Fig. 4e–g). In contrast, when we used dCas9 to target the Pfam library to endogenous genes, we found 68 hits targeting CD2 with sg717, which systematically led to stronger activation than any other sgRNA targeting CD2, CD20 or CD28 (Fig. 2c and Extended Data Fig. 4h–l). Only 19% of rTetR-activator hits at minCMV recurred as dCas9 hits at CD2 (for example, ZNF473 KRAB and SMARCA2 QLQ); conversely, 89% of domains capable of activating CD2 were not hits in the rTetR- and reporter-based screens, including 24 HLH domains (for example, NGN2 and HAND2). The second strongest HLH activator domain was from HAND2, consistent with a recent report that full-length HAND2 protein can be an activator in a recruitment assay³³. Indeed, 77% of the HLH activator domain hits were from proteins that were also activators in that study ($P = 3.1 \times 10^{-3}$, Fisher's exact test; Extended Data Fig. 4m). Meanwhile, the two MYB LMSTEN domains were only activators when recruited with rTetR at reporters. Using the larger CRTF tiling library, dCas9 recruitment to CD2 identified 19 activator domains that were not hits with rTetR at minCMV. In total, 42% of the CD2-only activators again overlapped with HLH domains (for example, in MYOD1, PTF1A, MAX and ASCL5), while others were new activators that did not overlap any domain annotation (for example, PBRM1; Extended Data Fig. 5a–d). A notably strong shared activator hit with both rTetR at minCMV and dCas9 at CD2 was the DUX4 C-terminus, which interacts with histone acetyltransferase P300 (ref. 35). Therefore, rTetR recruitment to different minimal promoters identified similar activators, while dCas9 recruitment to an endogenous gene identified a largely distinct set of activators including HLH domains (Supplementary Tables 1–3).

We then investigated the cell-type dependence of different activators by comparing their effects in HEK293T versus K562 cells when recruited by rTetR at the minCMV reporter. In general, we found much greater differences between activators targeted to the same promoter in different cell types than when targeting distinct minimal promoters within the same cell type. Overall, only 19% of activators in K562

cells were shared across the cell types, including strong activators like FOXO-TADs, NCOAs and the ZNF473 KRAB domain (Fig. 2d,e and Supplementary Fig. 1a). Meanwhile, several WW, SRC Homology 3 (SH3), plant homeodomain (PHD) and the poorly characterized SMARCA2/SMARCA4 QLQ domains^{36,37} were much stronger activators in K562 cells, despite being similarly expressed in the two cell types as measured by FLAG staining (Fig. 2e, Extended Data Fig. 3c and Supplementary Fig. 1a–d). We hypothesized that one source of cell-type dependence of an effector could arise from competition for coactivators with its endogenous copy, and found that knocking down the endogenous SMARCA2/SMARCA4 resulted in increased activation by the recruited SMARCA2 QLQ (from 1.3% to 23.7% of HEK293T cells ON; Supplementary Fig. 1e–i).

Overall, while we cannot rule out all technical reasons that these activators are stronger in K562 cells (for example, related to the reporter chromatin state or rTetR), these results show that some activators function very differently across cell types (for example, KIBRA WW activates 91% of K562 versus 3% of HEK293T cells) while others are more consistent (for example, FOXO3-TAD activates 93% of K562 versus 87% of HEK293T cells; Supplementary Fig. 1a,b).

Repressor function across targets and cell types

We next tested repressor promoter-specificity using rTetR to target pEF1 α , PGK and UbC promoters in K562 cells. We found repression scores at the moderate strength PGK and UbC were highly correlated with each other, largely due to signal from KRAB repressors ($R^2 = 0.74$ for $n = 2,718$ well-expressed Pfam domains; Supplementary Fig. 2a), whereas the stronger pEF1 α was more silenced by weaker repressors such as HOX homeodomains (Supplementary Fig. 2b,c). We previously observed that HOX homeodomain repression strength correlated with the presence of an RKRR motif and positive charge in the homeodomain N-terminal region³², so we recruited motif- and charge-modifying HOX mutants to the pEF1 α reporter and confirmed they contribute to repression (Supplementary Fig. 2d,e). In sum, the similarly strong PGK and UbC reporters responded similarly to repressors, whereas the stronger pEF1 α reporter uniquely identified weak non-KRAB repressors and thus could be used to characterize their sequence dependencies.

To identify repressors of endogenous genes, we used dCas9 recruitment. When targeting the pEF1 α reporter with dCas9, we found fewer hits than with rTetR (possibly due to lower recruitment copy number), and all the hits were KRAB repressors (Supplementary Fig. 2f–h). In contrast, targeting dCas9 to the endogenous CD43, we found that in addition to shared hits including KRAB domains and a zinc finger from the IRF-2BP1 corepressor protein³⁸, a distinct set of domains that were not hits in the rTetR screens showed repressor activity, including the NuRD-interacting methyl-binding protein domains such as the P66_CC domain from P66A and the MBDA domain from MBD2 (which bind one another³⁹) (Fig. 2f and Supplementary Fig. 2i–k). Relatedly, an MBD2B repressor was previously shown to silence a reporter in HEK293T cells with dCas9 recruitment⁴⁰. Another repressor hit unique to targeting CD43 with dCas9 was the SAM1 domain from the putative polycomb protein L3MBTL4 (whose analog is involved in silencing in *Drosophila* cells⁴¹). MBDA, P66_CC, IRF-2BP1 and SAM1 silencing of CD43 depended on which sgRNA was used, while KRAB repressors were efficient with either of two sgRNAs (Supplementary Fig. 2i,j). Meanwhile, chromo and chromoshadow domains were only hit with rTetR.

dCas9 recruitment of the CRTF tiling library to the CD43 promoter confirmed these results, with the strongest shared repressors being KRAB domains, and additionally revealing 274 repressor domains that were not hits at pEF1 α , including from additional methyl-binding domain proteins (Extended Data Fig. 6a–c). This larger library also uncovered repressors in the unannotated regions of proteins, including CDCA7L, which is known to be a repressive protein⁴². In total, 78% of CD43 repressor domains were also hits in the rTetR screens, and

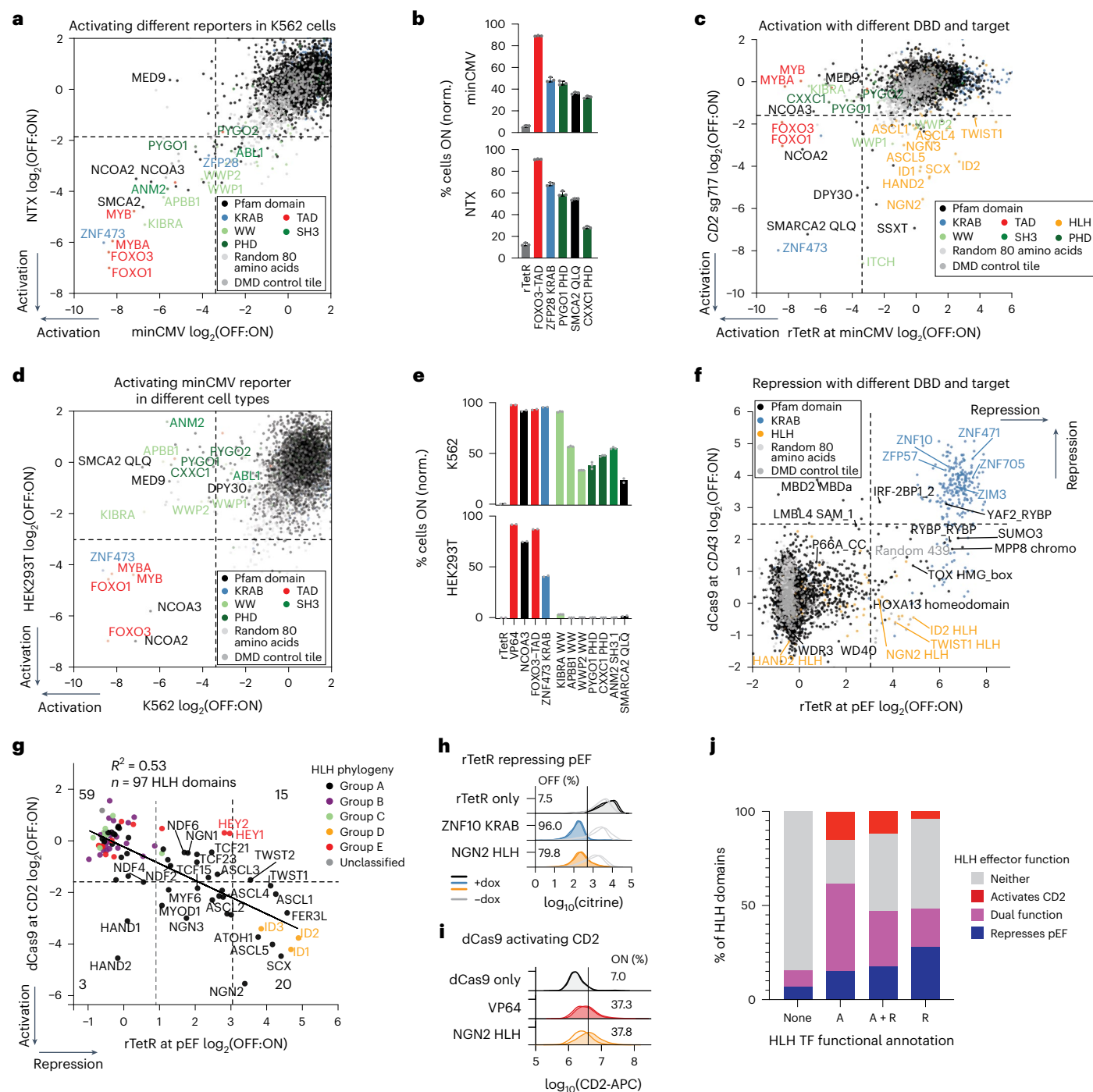


Fig. 2 | Context-dependent transcriptional effectors. **a**, HT-recruit with rTetR targeting two minimal promoters, minCMV and NTX, in K562 cells ($n = 2$ biological replicates). Dashed lines show hit thresholds at 2 s.d. above the median of the poorly expressed domains (same in **c**, **d** and **f**). **b**, Validation of activator domains across minimal promoter reporters in K562 cells. Individual rTetR-activator fusions or the rTetR-only negative control were delivered by lentivirus, and, after selection, cells were treated with 1,000 ng ml⁻¹ doxycycline for 2 days to induce reporter activation. The percentage of cells activated was measured by flow cytometry for the citrine reporter, after gating for delivery with mCherry. Bars show the average percentage of cells ON normalized to no-doxycycline control; error bars are s.d. ($n = 3$ infection replicates shown as dots). **c**, HT-recruit with dCas9 targeting endogenous gene *CD2* with sg717 compared with rTetR targeting the minCMV reporter in K562 cells ($n = 2$ biological replicates). **d**, HT-recruit with rTetR targeting the minCMV reporter in K562 and HEK293T cells ($n = 2$ biological replicates per cell type). **e**, Individual rTetR fusions were measured in minCMV reporter cells as in **b** ($n = 2-3$ infection

replicates shown as dots). **f**, HT-recruit with dCas9 targeting endogenous gene *CD43* with sg15 compared with rTetR targeting the pEF1 α reporter in K562 cells ($n = 2$ biological replicates). **g**, HT-recruit with dCas9 to activate *CD2* using sg717 compared with repression of pEF1 α promoter with rTetR in K562 cells, showing only the HLH domains within the Pfam library ($n = 2$ replicates per screen). Black line shows the best linear fit. The conservative hit threshold (black dashed line) was chosen to identify strong effectors. The gray dashed vertical line is equivalent to the strength of the weakest repressor that was individually validated (Methods). The HLH phylogenetic groups are shown in colored solid circles⁴⁴⁻⁴⁶. **h**, rTetR recruitment to the pEF1 α reporter in K562 cells with 6 days of treatment with 1,000 ng ml⁻¹ of doxycycline ($n = 2$ infection replicates shown as histograms). **i**, dCas9 recruitment to *CD2* in K562 cells ($n = 2$ sgRNAs, sg717 in darker and sg718 in lighter shade). **j**, Full-length HLH TFs were defined as not yet defined (none, $n = 45$), activators (A, $n = 13$), both (A + R, $n = 17$) or repressors (R, $n = 25$), in previous studies reviewed in ref. 46. Colors show the effector function of HLH domains from these TFs.

they overlapped similar annotations for SUMOylation, short linear interactions motifs and DBDs³⁴ (Extended Data Fig. 6d).

We then investigated the cell type-specificity of repressors using rTetR to target the Pfam library at pEF1α reporters. The HEK293T screen identified strong repressors that were in agreement across cell types, such that 96% of repressor hits in HEK293T cells were also hits in K562 cells (including >200 KRAB domains; Supplementary Fig. 3a–d). When using the moderately strong UbC promoter, we found fewer hits than with pEF1α, and again the majority were shared across cell types (for example, >130 KRAB domains; Supplementary Fig. 3e). Altogether, these results show that some repressors (for example, MDBa) strongly depend on recruitment context, while KRAB repressors are particularly robust across cell type, target and DBD contexts.

HLH effectors can activate or repress depending on the context

Effector domains are classified as activators or repressors; however, like the TFs that consist of these domains⁹, these effectors may have a dual activator–repressor function³⁴ that could be context-dependent. When comparing most activatable and repressible contexts, we saw no overlap in hits from the Pfam library; however, when considering the most sensitive repressible context (rTetR at pEF1α in K562 cells), we found some activators could repress (Supplementary Fig. 4a–d). Certain minCMV strong activators either silence a percentage of the pEF1α cells (that is, bifurcate the population) or super-activate it in a cell type-dependent manner (Supplementary Fig. 4e,f).

In addition, the large HLH family, which includes TFs shown to function either as an activator or repressor depending on context in yeast⁴³, stood out as a major source of dual-functioning effectors. Specifically, many of the same HLH effectors that activate CD2 when recruited with dCas9 also repressed pEF1α when recruited with rTetR with correlated strength ($R^2 = 0.53$, $n = 97$ HLH domains; Fig. 2g–i and Extended Data Fig. 7a). Furthermore, we found that effector function is enriched within certain HLH phylogenetic groups^{44–46}; for example, HAND HLHs from group A are only activators, HEY HLHs from group E are only repressive, ID HLHs from group D are dual-functioning and other subfamilies (for example, HES from group E and all of group B) did neither in the contexts tested here (Fig. 2g and Extended Data Fig. 7b). These HLH effector functions partially corresponded with annotations from previous studies of the HLH TFs as activators, repressors or both (Fig. 2j)⁴⁶. These results were confirmed with the tiling library, where 74% of proteins with a dual-function tile that activates CD2 (but not minCMV) and represses pEF1α were again HLH proteins. The hit tiles overlapped the HLH heterodimerization portion of their bHLH domains, and deletion experiments showed that this C-terminal portion was required for both activation and repression with a dual-functioning HLH (Extended Data Fig. 7c–g)³⁴.

Overall, while some effectors were highly context-dependent (for example, HLH and WW domains), we noticed many of the strongest effectors were consistent across contexts (for example, KRAB repressors), which suggests they would improve synthetic tools for manipulating transcription.

Efficient CRISPRi with ZNF705/ZNF471 KRAB repressors

We set out to identify repressors that are robust across contexts using the Pfam library because it was screened against the largest number of contexts. We performed an additional screen with this library at the *GATA1* locus to compare repressors at an enhancer (Supplementary Fig. 5a). *GATA1* is an essential gene, so we used the growth phenotype associated with targeting dCas9-repressors to its enhancer (eGATA1) as a proxy for repression strength^{14,47}. The KRAB domain found in ZNF705B/ZNF705D/ZNF705F (here called ZNF705) and ZNF471 were the strongest hit Pfam domains with the eGATA1 sgRNA, and they did not have growth effects with a control safe-targeting sgRNA (Supplementary Fig. 5b and Supplementary Table 1). Meanwhile, some domains did show growth effects with the safe-targeting sgRNA, suggesting

their expression is toxic. The most toxic were the cyclin-dependent kinase inhibitor domains from CDKN1A/CDKNB, which inhibit cell cycle progression by binding to CDK2 (ref. 48). The random sequences were substantially more toxic than the Pfam domains or DMD tile controls (Supplementary Fig. 5c,d), which could be due to random sequence misfolding⁴⁹. We confirmed these toxic domains were not confounding repressor hits when targeting the nonessential gene, *CD43* (Supplementary Fig. 5e).

To identify strong repressors that function well across many different contexts, we focused on the KRAB domains, which were commonly top hits. Furthermore, despite some effort, we could not identify more efficient non-KRAB repressors (Supplementary Note 1 and Supplementary Fig. 6a–l), consistent with KRAB repressors being most potent across contexts in the screens. Previously, using a deep mutational scan of the ZNF10 KRAB, we identified a mutant in the N-terminal KRAB domain region, WSR7EEE, that provides increased expression in cells and silencing strength when fused to rTetR³². We found the same is true when this mutant is fused to dCas9 (Fig. 3a,b and Extended Data Fig. 8a,b). This enhanced mutant KRAB specifically silences endogenous genes via heterochromatinization (as marked by H3K9me3), similar to reported data for the CRISPRi tools that use the WT KRAB from ZNF10 (refs. 5,50; Fig. 3c). We previously found that nearly all KRAB paralogs that repress in HT-recruit also interact with KAP1 as measured by mass spectrometry and colocalize with H3K9me3 by chromatin immunoprecipitation followed by sequencing (ChIP-seq), so we expect they also recruit H3K9me3 (ref. 32). Therefore, we used this enhanced ZNF10 KRAB (WSR7EEE) as a benchmark when determining how well the KRAB repressor paralogs work.

We ranked 323 KRAB domains by their scores across all of the repressor screens and found ZNF705 KRAB ranked 18 and ZNF471 KRAB ranked 1 (Fig. 3d). In agreement with a recent finding from a screen of 57 KRABs, our approach also identified ZIM3 KRAB (13) as a stronger repressor than ZNF10 KRAB (166)^{51,52}. Further supporting their context-robustness and reproducibility, we found ZNF705, ZNF471 and ZIM3 KRAB were hits in both replicates of 8, 7 and 6 of the original 8 repressive contexts, respectively (Fig. 1e and Supplementary Table 1). Directly comparing HT-recruit scores across contexts suggested that these top KRAB domains were stronger than ZNF10 KRAB but not meaningfully different from one another ($n = 12$ contexts; Extended Data Fig. 8c).

We set out to validate the top-ranked KRAB domains across different target contexts compared to the strong ZNF10 KRAB (WSR7EEE). Encouragingly, the KRAB domains from ZNF705 and ZNF471 both outperform the ZNF10 KRAB (WSR7EEE) at silencing CD43 (Fig. 3e). To test additional targets, we performed CRISPRi screens with a library including 405 sgRNAs targeting promoters of 37 essential genes (Extended Data Fig. 8d,e) and found that the ZNF705 KRAB consistently provides ~1.5× greater effect sizes across a range of ZNF10 baseline effects and sgRNA distances from the TSS (Fig. 3f and Extended Data Fig. 8d–g). We also found that ZNF705 KRAB consistently exhibits stronger effects at candidate *cis*-regulatory elements (Fig. 3g). These KRABs also completely silence a reporter in HEK293T cells (Extended Data Fig. 8h,i). Furthermore, ZNF705 KRAB more completely silenced an endogenous gene than ZNF10 KRAB across a population of THP-1 monocytes when stably delivered by lentivirus ($n = 2$ sgRNAs; Fig. 3h and Extended Data Fig. 8j). ZNF705 matched ZNF10 KRAB when transfected into iPSCs, which could be a saturated condition (Extended Data Fig. 8k,l). We note that the enhancing ZNF10 (WSR7EEE) mutation is in a unique N-terminal region that is spliced onto the ZNF10 KRAB A-box from a distinct exon not found in other KRAB genes. So, we could not transfer that same mutation to further enhance ZNF705/ZNF471/ZIM3.

To assess the KRAB repressors in a different DBD context, we used an engineered form of dAsCas12a (dEnAsCas12a; 1,308 amino acids)^{53,54}. dCas12a is weaker than dCas9 for recruiting repressors, so there is more need to improve its efficiency¹⁸. Fusing ZNF705 KRAB to dCas12a

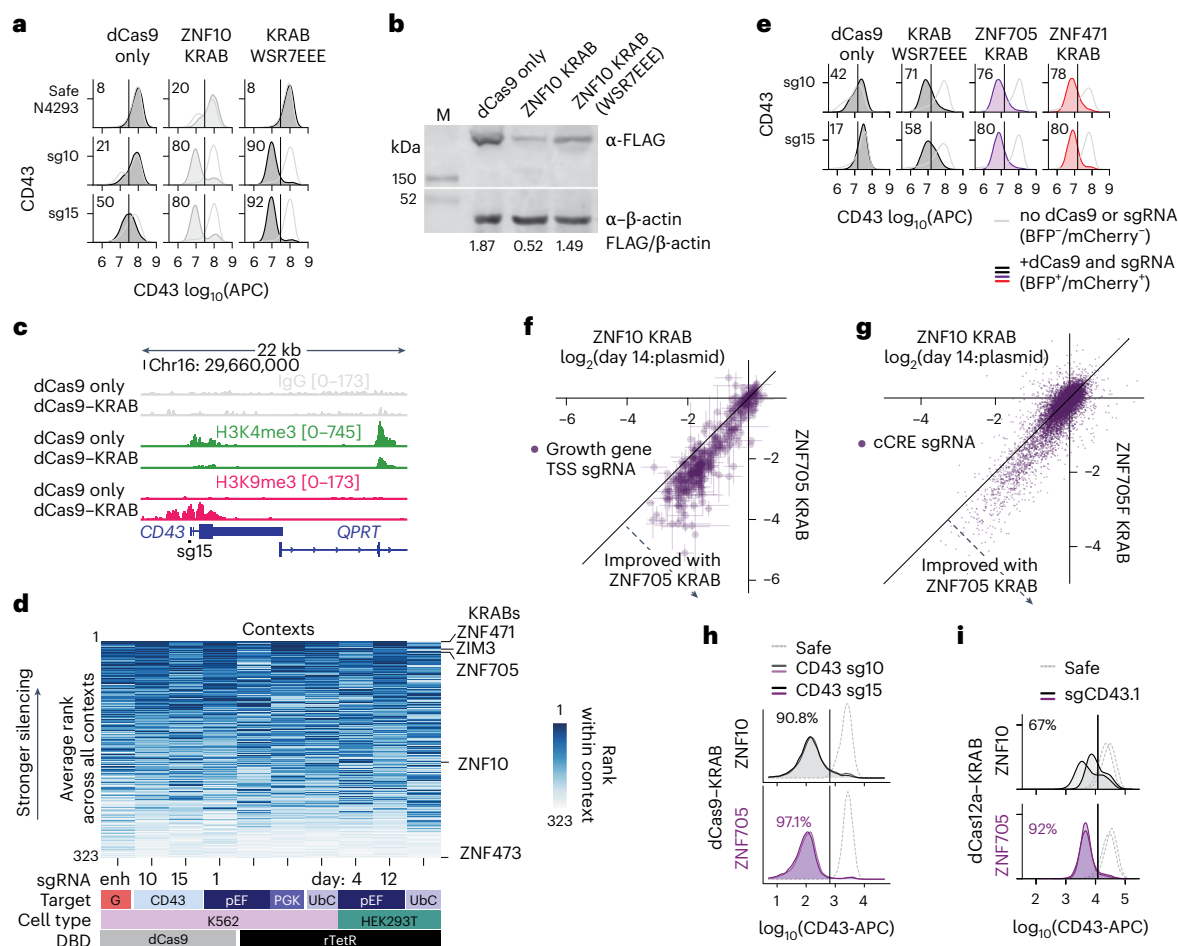


Fig. 3 | Improved CRISPRi using ZNF705 and ZNF471 KRABs. **a**, dCas9 recruitment in K562 cells. sgRNAs were delivered by lentivirus, followed by dCas9 constructs, and the cells were stained 7 days after dCas9 infection. The shaded histogram shows the cells after gating for both the dCas9 (BFP) and the sgRNA (mCherry), and their percentage of cells OFF is shown. The unshaded histogram shows the cells from the same sample that express neither and serve as an internal control ($n = 1$ infection replicate). **b**, Western blot of dCas9 only, ZNF10 KRAB and ZNF10 KRAB (WSR7EEE) fusions ($n = 1$ biological replicate). dCas9 and KRAB fusions were tagged with 3xFLAG. The band intensity ratio is below. **c**, Chromatin modifications mapped by CUT&RUN after dCas9 recruitment of ZNF10 KRAB (WSR7EEE) using sg15. Stable cell lines expressing the dCas9 fusion and the sgRNA were selected with antibiotics and fluorescent cell sorting before chromatin was analyzed. **d**, KRAB domains ranked by repression in HT-recruit screens across the target, cell type and DBD contexts (a higher average $\log_2(\text{OFF:ON})$ from two biological replicates is better ranked). The 323/336 KRAB domains that had no missing data across contexts are included. The red target labeled G shows the growth-based screen at the GATA1 enhancer. The HEK293T pEF screen has two time points as follows: a silencing measurement at 4 days after doxycycline addition and a memory measurement at day 12 (8 days after doxycycline removal). **e**, sgRNAs were delivered by lentivirus, followed by dCas9

constructs, and the cells were stained 9 days after dCas9 infection ($n = 1$). **f**, CRISPRi growth screen to compare KRAB repressors. A sgRNA library targeting 37 essential gene promoters was delivered into K562 cell lines that stably express either dCas9–KRAB. Cells were passaged for 14 days, and then the guides were sequenced to measure fitness effects (change from the original plasmid pool to the final day 14 genomic DNA). Greater depletion is a measure of stronger silencing of the essential genes. Each dot shows the average effect for a sgRNA, and the error bars show the s.d. from two screen replicates ($n = 405$ sgRNAs). The diagonal line represents the identity between KRAB domains. **g**, Results are shown similarly for the sgRNAs in the library that target cCREs of growth genes ($n = 10,155$ sgRNAs). **h**, sgRNAs were delivered to THP-1 monocytes by lentivirus, followed by dCas9 fusions. Eight days later, cells were stained. Gating and shading as in **a** ($n = 2$ targeting sgRNAs shown as histograms). **i**, dCas12a recruitment in K562 cells. dCas12a–KRAB fusions and then guide RNAs were delivered by lentivirus. Nine days after guide RNA infection, cells were stained. Data are gated for guide RNA expression (mCherry) and dCas12a expression (hemagglutinin (HA)-tag stain). Infection replicates are shown as separate histograms ($n = 2$ biological replicates shown as histograms). cCREs, candidate *cis*-regulatory elements.

resulted in a mean $1.5\times$ more complete silencing of a population of cells than ZNF10 KRAB across multiple targets (Fig. 3i and Extended Data Fig. 8m). Encouragingly, a 99% identical ZNF705 family KRAB (from ZNF705E) was ranked 12 of $>1,000$ repressor hit tiles recruited in three contexts with the larger tiling library, further establishing ZNF705 KRAB as a particularly strong repressor (Extended Data Fig. 8n,o). Other highly ranked KRABs likely perform similarly well. Concordant with its low toxicity and previous RNA-sequencing (RNA-seq) results with ZNF10 KRAB^{55,56}, stable overexpression of dCas9–ZNF705 KRAB did not perturb the baseline transcriptome relative to a dCas9-only control (0.01% transcripts changed $>4\times$ by RNA-seq; Extended Data Fig. 8p).

Compact human activator combinations for lentiviral CRISPRa
We next ranked activators from HT-recruit measurements performed across the target, DBD and cell-type contexts and selected context-robust domains that were a hit in at least five activation screens (Fig. 4a). ZNF473 KRAB (Z) is an acidic domain that was a strong hit with both dCas9 recruitment to CD2 and most rTetR contexts. Meanwhile, NCOA3 (N) and FOXO3–TAD (F), which bind p300 (refs. 57–59), were rTetR hits and stronger than ZNF473 in HEK293T cells. Previous ChIP experiments showed that the full-length proteins from which we sourced these effectors are associated with active chromatin marks—NCOA3 is enriched near H3K27ac and H3K4me1 chromatin

modifications⁶⁰, FOXO3 colocalizes with RNA polymerase II and H3K27ac^{61,62} and ZNF473 is enriched at H3K27ac peaks³². We found that these domains were well-expressed across DBDs and cell types and there was no toxicity associated with their expression (Extended Data Fig. 3d and Supplementary Fig. 5e). We had previously found that the trimmed Pfam-annotated domains (Z = 44 amino acids, N = 48 amino acids and F = 41 amino acids) for these three activators performed equally to the 80 amino acid sequence centered on the domain with rTetR³², so we used the trimmed sequences to build activator tools (Fig. 4b). This trimmed FOXO3 sequence was within the 11th activator tile in the larger library ranked by the average of the minCMV and CD2 screens (Supplementary Table 2).

We fused these trimmed activators, or two commonly used synthetic activators (VP64 and VPR)^{63,64}, individually to dCas9 in a lentiviral vector with BFP as a delivery marker. Given previous observations of synergistic activation from concatenated activators (including VP64 compared to VP16; VPR compared to its components VP64, p65 and Rta; and the synergistic activation mediator (SAM), which combines VP64 with a >450 amino acids MCP-p65-HSF1 component recruited via MS2 stem-loops in the sgRNA)^{4,64–68}, we reasoned that fusing N, F and Z could improve activation. Indeed, when we combined any of these domains together as bipartite activators, we observed synergistic activation of the *CD2* gene (Fig. 4c). Further fusion of all three domains together to create the tripartite activator, NFZ, led to even stronger activation (Fig. 4c).

At 145 amino acids long, NFZ is more compact than the tripartite 523 amino acid VPR and does not use viral components. We found that NFZ was more efficiently delivered by lentivirus, more efficiently generated stable cell lines after blasticidin selection and provided a similar or higher level of activation across three target genes (Fig. 4c–f and Extended Data Fig. 9a–d). Furthermore, we could use lentivirus to deliver NFZ, but not VPR, to J774 mouse macrophages, which are more difficult to infect than K562 cells, and successfully select a stable line in which we could activate endogenous genes (Fig. 4g and Extended Data Fig. 9e,f). NFZ was also a potent activator in THP-1 monocytes and iPSCs (Fig. 4h,i). Additionally, by changing the N, F and Z domain orientations, we identified NZF as a moderately stronger activator with lentiviral delivery efficiencies between VPR and NFZ (Fig. 4f and Extended Data Fig. 9a,b,g).

NFZ may be more deliverable by lentivirus than VPR due to differences in transduction efficiency and/or cell toxicity related to the

expression of VPR. While VPR is the largest effector, the lentiviral insert size (between central polypurine tract (cPPT) and woodchuck hepatitis virus post-transcriptional regulatory element (WPRE)) is 8.5 kb, which does not exceed packaging limits⁶⁹. Meanwhile, transcriptional squelching is associated with VP16-derived activators^{70,71}, and VPR can be toxic in cell lines⁷², which could affect the packaging cells and/or transduced cells. When using transient plasmid delivery, we observed similar or better activation with VPR than NFZ (Extended Data Fig. 9h,i), despite observing lower expression of VPR transcripts (Extended Data Fig. 9j,k; consistent with others' observations of low dCas9-VPR expression^{73,74}), suggesting NFZ is specifically advantageous for viral delivery.

Notably, the tripartite fusions were strong activators even with dCas9 targeting CD20, CD28, CD2 (with its sgRNAs beyond sg717) or the minCMV reporter, which were all contexts with poor activation from nearly any single domain in the Pfam library (Fig. 4c,f, Extended Data Fig. 9g–i, l–n and Supplementary Table 1). We found no further improvement by fusing a fourth domain, QLQ, or using homotypic combinations (Extended Data Fig. 9l–o and Supplementary Note 2).

One of the most potent currently used CRISPRa tools is the larger SAM system⁶⁶. SAM consists of the following three components: dCas9–VP64, a sgRNA engineered to contain the MS2 stem-loop and a 472 amino acid helper protein MPH (MS2-binding protein fused to P65 and HSF1 human activators; Supplementary Fig. 7a). We found that we could replace the viral VP64 in SAM with human NFZ/NZF and activate endogenous CD2 with comparable efficiency (Supplementary Fig. 7b–g and Supplementary Note 3). We also found the two-component dCas9–NFZ easier to deliver than the three-component SAM (which required an additional helper protein virus, thus resulting in fewer cells co-infected with all components), but that SAM can be more potent if successfully delivered.

Fusing compact activator NFZ to smaller CRISPR DBDs

dCas9 is a relatively large DBD (1,368 amino acids), and it is difficult to multiplex with multiple sgRNAs. To make a CRISPRa system that can be multiplexed, we ported NFZ to dEnAsCas12a (1,308 amino acids)^{54,75} and found that NFZ activated CD2 approximately tenfold more strongly than VP64 (Fig. 4e,j and Supplementary Fig. 8a,b). As with dCas9, we could not establish a stable dCas12a–VPR cell line using lentivirus (7.6 kb) and blasticidin selection, but we were successful with dCas12a–NFZ (Supplementary Fig. 8c,d).

Fig. 4 | Compact NFZ activator improves CRISPRa systems in different cell types.

a, HT-recruit scores for activators that were hit in ≥ 5 samples across the target, cell type and DBD contexts ($n = 2$ replicates per rTetR screen and $n = 1–2$ replicates per sgRNA for dCas9 screens shown as columns). The rows are clustered in an unbiased manner. Labels for NCOA3 (1,045–1,092; N), FOXO3 FOXO–TAD (604–644; F) and ZNF473 KRAB (5–48; Z) are colored. **b**, Size of N, Z and F human activator domains and combinations compared to viral activators. **c**, The effectors were recruited with dCas9 in K562 cells. sgRNAs were delivered by lentivirus, and cells were selected with puromycin. Then dCas9 fusions were delivered by $10\times$ concentrated lentivirus. Four days after dCas9 delivery, cells were stained and measured by flow cytometry ($n = 2$ infection replicates shown as histograms). Left, CD2 expression after gating cells for sgRNA delivery (GFP). Middle, the average percentage of cells with dCas9 delivery (BFP) is written in blue. Right, CD2 expression after gating for both sgRNA and dCas9 delivery. The darker histogram is CD2 sg717, and the lighter shade is sg718. **d**, Comparison of activation and dCas9-activator delivery (BFP) after gating for sg717 delivery (GFP⁺). Color shows the smoothed density of flow cytometry data. The percentage of BFP⁺ cells is labeled ($n = 2$ infection replicates). **e**, Schematics of CRISPR activator systems using the NFZ tripartite activator fused to dCas9, dEnAsCas12a or dCasMINI_V4. **f**, dCas9 activators were delivered by $5\times$ concentrated lentivirus into stable sgRNA-expressing cell lines; 5 days later, blasticidin selection was started, and then 4 days later, cells were stained. Top, the percentage of cells ON (APC) is shown after gating for the sgRNA (GFP⁺) and dCas9 (BFP⁺). All of the VPR samples had <250 BFP⁺ cells counted

(associated with poor infection and survival of blasticidin selection), while all other samples had >6k BFP⁺ cells (mean = 53k). Bottom, the percentage of BFP⁺ cells is shown ($n = 5–7$ sgRNAs per effector). The CD28 sgRNAs infected with VPR poorly survived blasticidin selection, and <300 cells total could be counted, so they are not shown. **g**, dCas9–NFZ was stably delivered to J774 mouse macrophages by lentivirus and selected with blasticidin. Activation of endogenous Gpr84 or Actc1 measured by qPCR after lentiviral sgRNA delivery ($n = 3$ qPCR replicates). NT sgRNAs are negative controls. **h**, THP-1 monocytes stably expressing the sgRNAs were generated using lentiviral delivery, followed by infection with dCas9-activator lentiviruses. Cells were stained for CD2 expression 8 days later. Gated for sgRNA delivery (GFP) and dCas9 (BFP; $n = 1$ infection per sgRNA). **i**, iPSCs were transfected with dCas9 activator and sgRNA plasmids and then stained 2 days later. Gated for sgRNA delivery (GFP) and dCas9 (BFP; $n = 3$ transfection replicates shown as histograms). The percentage of cells ON with the gene-targeting sgRNA, using the threshold shown by the black vertical line, is shown. **j**, dEnAsCas12a activators on a marker-less plasmid were co-electroporated into K562 cells with a gRNA-mCherry expression plasmid. After 3 days, cells were stained. Gated for high mCherry ($n = 2$ replicates). **k**, dCasMINI activators on a plasmid with an mCherry marker were cotransfected into HEK293T GFP reporter cells with a gRNA-BFP expression plasmid. Two days later, GFP reporter activation was measured by flow cytometry with gates for BFP⁺/mCherry⁺ cells ($n = 6$ transfection replicates shown as dots, and bar shows mean). Np NLS, nucleoplasmic nuclear localization signal; NT, nontargeting.

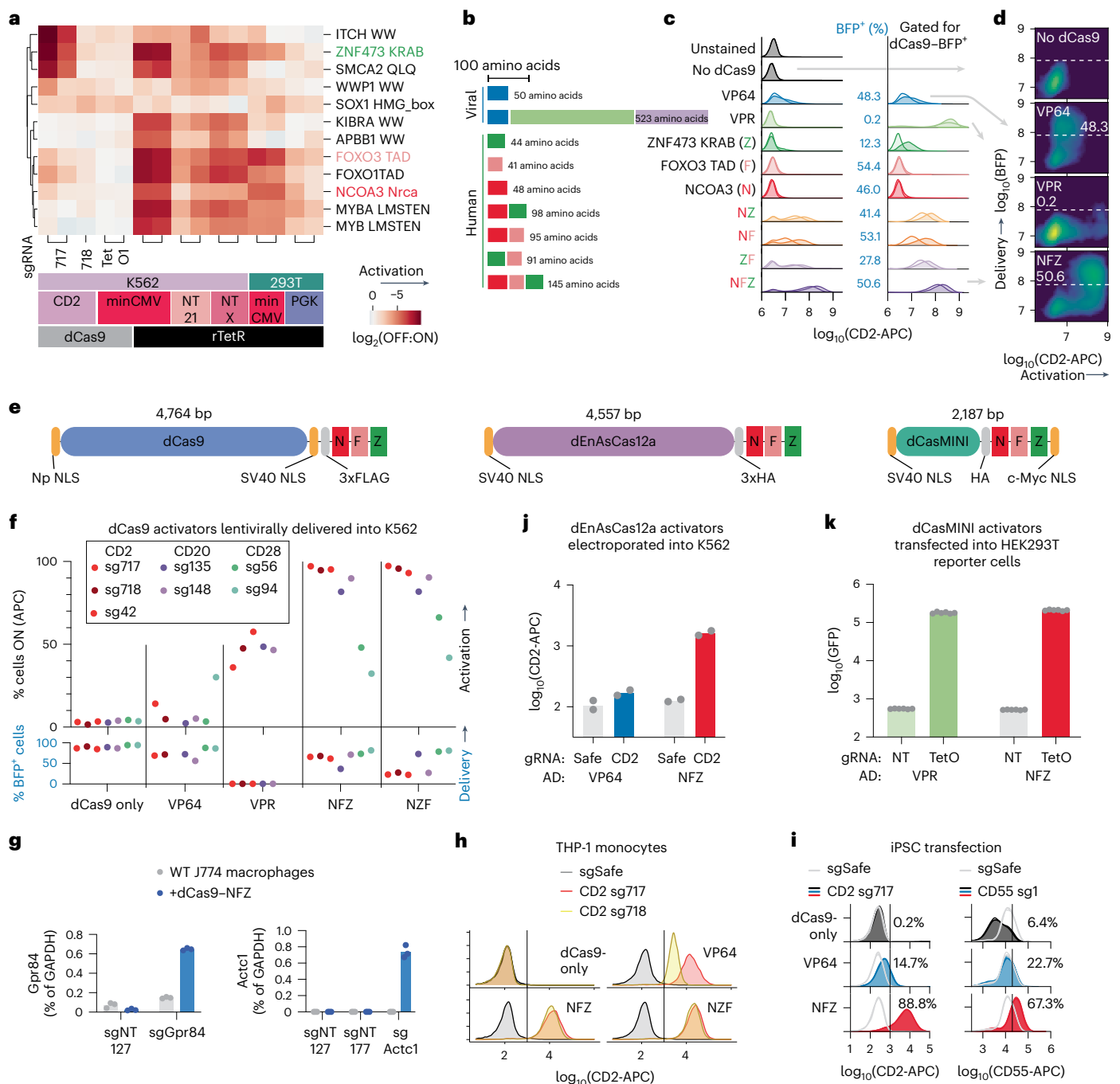
Recently, a very compact dCas12fDBD called dCasMINI (529 amino acids) was engineered as a CRISPRa system in human cells⁷⁶. We found that dCasMINI-NFZ (2.2 kb) strongly activated HEK293T reporter cells (Fig. 4e,k and Supplementary Fig. 9a–e).

Inducible expression with the compact activator NFZ

Another useful application of compact human activators is engineering circuits that tunably control transgene expression using a synthetic TF that conditionally activates a promoter in the presence of a small molecule⁷⁷. Previously, the zinc finger-homeodomain DBD (ZFHD1)⁷⁸ was fused to FK506 binding protein (FKBP)-dimerization domains that recruit HSF1 and p65 activators in the presence of rapamycin⁷⁹. We designed a >1 kb smaller version of this design using NFZ (Fig. 5a and Supplementary Fig. 10a). Upon transfection, we observed rapamycin-dependent reporter transgene inducibility and low levels

of leakiness with NFZ relative to P65 + HSF1 (Fig. 5b and Supplementary Fig. 10b–d). Next, we used ZFHD1–NFZ to inducibly express the human hepatocyte growth factor (HGF) gene (2.2 kb)^{80–84} and observed a rapamycin dose-dependent increase in secreted HGF protein, with minimal leakiness in untreated cells (Fig. 5c). This compact circuit could enable longer proteins (such as HGF) to be inducibly expressed from limited-size viral vectors like adeno-associated virus (AAV).

Inducible gene expression circuits can also expand the therapeutic potential of chimeric antigen receptor (CAR) T cells by providing control over CAR expression. Such a circuit, built using synthetic zinc finger transcription regulators (synZIFTR), was recently shown to be effective for dose- and time-dependent control of CARs in therapeutically relevant primary T cells both in vitro and in mouse models of cancer⁸⁵. To minimize immunogenicity, synZIFTRs use a human activator domain. Here by replacing the previously used p65 activator with NFZ,



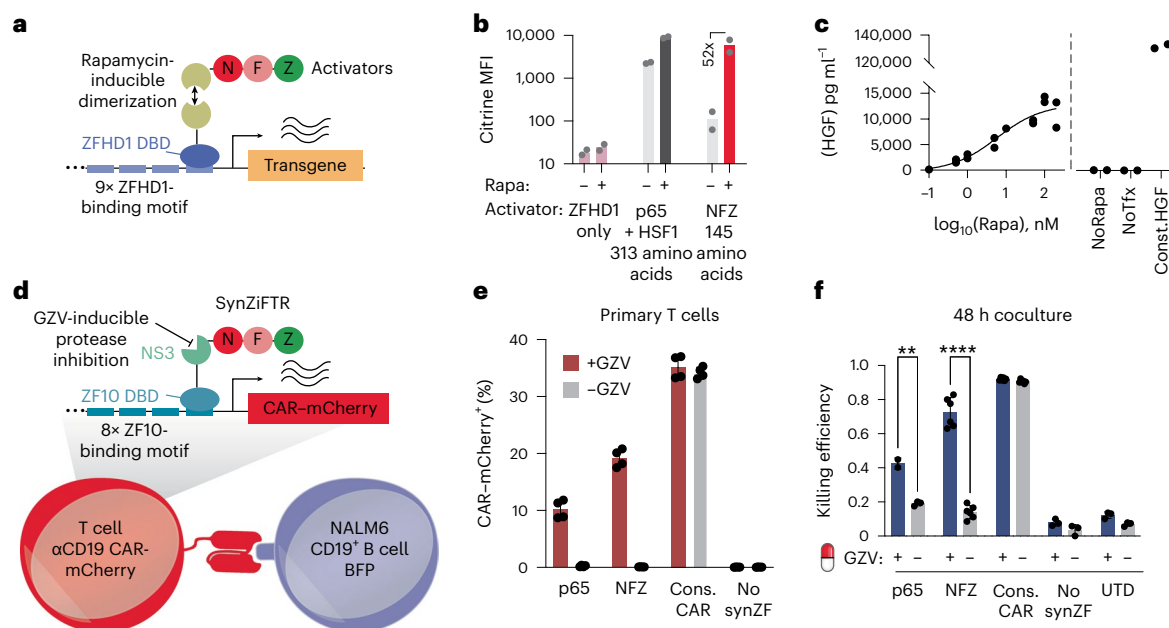


Fig. 5 | Compact NFZ activator improves therapeutically relevant inducible systems. **a**, Schematic representation of the minimized rapamycin-inducible transcription cassette using the ZFHD1 DBD fused to NFZ. **b**, Rapamycin-inducible expression of citrine with ZFHD1 recruitment of NFZ or a control activator combination, p65 + HSF1. In total, 3 μ g of plasmids were transfected into HEK293T cells; 1 day later, 10 nM rapamycin was added, and then 2 d later, citrine MFI was measured by flow cytometry ($n = 2$ transfection replicates shown as dots, and bar shows mean). **c**, Rapamycin-inducible expression of HGF with ZFHD1 recruitment of NFZ, after transfection in HEK293T cells. Secreted HGF protein concentrations were measured in the cell culture supernatant by ELISA after 2 days of rapamycin treatment with varied doses. Transfection of no plasmid and a constitutive pEF1 α -HGF plasmid served as a negative and positive control, respectively ($n = 2$ transfection replicates; the curve shows nonlinear regression fit to dose-response data). **d**, Schematic representation of synZiFTR circuit for drug-regulated control of CAR T cells. SynZiFTR uses the FDA-approved drug Grazoprevir (GZV) to inducibly inhibit NS3 protease-mediated self-cleavage of

an engineered zinc finger activator. Upon induction, the activator is recruited to synthetic ZF10-binding motifs⁸⁵ to activate an anti-CD19 CAR fused to a mCherry reporter. Bottom illustration shows a synZiFTR CAR T cell targeting a CD19⁺ cancer cell. **e**, Efficiency of activating CAR-mCherry as measured using the vertical threshold shown in Extended Data Fig. 10c ($n = 1-2$ infection replicates measured with two technical replicates each shown as dots, and bar shows mean and error bars show s.d.). ‘Cons. CAR’ control is the constitutive spleen focus-forming virus (SFFV) promoter, and ‘no synZF’ control only contains the CAR-mCherry transgene. **f**, After 72 h of induction with GZV, synZiFTR T cells were cocultured in the continuous presence of GZV for 48 h with a CD19⁺ NALM6 precursor leukemia B cell line engineered to express BFP. Killing efficiency was estimated by measuring the reduction in BFP⁺ cells compared to a NALM6-only negative control by flow cytometry ($n = 1-2$ infection replicates measured with three technical replicates each shown as dots, bars show mean and error bars show s.d.). ** $P < 0.005$ and **** $P < 0.00005$, by two-sided unpaired t test. UTD, untransduced; MFI, mean fluorescence intensity.

we found that we could increase induced CAR expression in primary T cells and correspondingly increase pro-inflammatory antitumor cytokine secretion and CAR-dependent cancer cell-killing efficiency (Fig. 5d–f and Extended Data Fig. 10a–h).

Discussion

A better understanding of transcriptional effector context-specificity is needed to improve strategies for manipulating transcription. Our systematic effector measurements across cell type, genomic target and DBD contexts revealed both context-dependent effectors (for example, WW and HLH domains) and context-independent effectors that can improve transcriptional perturbation tools (for example, the tripartite NFZ activator and the ZNF705 KRAB repressor).

For activation, NFZ may overcome delivery challenges associated with the size, number of components or reported toxicity of expressing VPR and SAM^{72,86–88} (Fig. 4d). NFZ is a compact, nonviral and low-toxicity activator that was better delivered by lentivirus than VPR (likely due to lower impacts on packaging cell or recipient cell health) and than SAM (because it does not require an additional helper protein component). In some applications, the domain orientation NZF provides stronger activation, but NFZ showed better lentiviral delivery and less leaky activation in an inducible circuit. Furthermore, NFZ shows promise as a compact and stably deliverable activator that could be easily multiplexable with dCas12a if further optimized, potentially in combination with other recent improvements for that DBD^{75,89–91}. Overall, our data suggest that NFZ could be preferable over VPR or SAM in contexts

where the larger systems cannot be equally well-delivered and where human-derived activators are needed, such as synZiFTR CAR T cells. However, we caution that the domain junctions in NFZ could produce immunoreactive peptides and that NFZ may not improve delivery or avoid toxicity in all contexts. CRISPRi/CRISPRa expression alone, with certain effectors, can perturb baseline gene expression of some genes^{56,68}, in a manner that correlates with the effectors’ toxicity⁵⁵, and NFZ could still have this issue despite being less toxic than VPR.

For repression, we find that KRABs are the strongest family across contexts and that ZNF705, ZNF471, ZIM3 and several other top-ranked KRAB domains are all similarly strong, demonstrating that repressors substantially stronger than ZNF10 KRAB are abundant upon screening all 336 KRABs. This wealth of options of strong KRAB domains could be useful to the field of synthetic biology, which now does not need to rely on only ZIM3 or ZNF10. Further studies could determine whether there are meaningful differences between these top KRABs or whether non-KRAB repressors from this resource outperform these top KRABs singly or in combinations in certain contexts (for example, target sites, cell types or recruiter fusion protein configurations).

Furthermore, our larger tiling library both confirmed context effects we originally observed with fewer elements in the Pfam library (for example, in the HLH, KRAB and MBD families) and provides a valuable resource of transcriptional effectors, including from unannotated protein regions, that function on dCas9 at endogenous genes. These transcriptional repressors and activators, in addition to others^{32,34,40,51,74,92–96}, can now be combined with varied DBDs⁸⁵ to

enable gene regulation tools that are more compact, humanized and deliverable.

Looking beyond the context-robust effectors, this and other recent work^{97,98} describe the extent of context dependence across effectors. Overall, for both activators and repressors, differences across cell types were more likely to be changes in magnitude, whereas differences across target and DBD were more likely to be complete changes from effective to ineffective (or even from activating to repressing). More specific trends can be found within domain families such as the target-dependent HLH domains or the cell-type-dependent WW and SH3 domains that interact with proline-rich motifs^{99,100} that are common in activators¹⁰¹. This resource of effector domains measured at endogenous sites provides useful starting points to explore mechanisms that explain which domains work at which loci. Integrating this effector function data with protein interaction maps could identify cofactors that help explain context dependence.

Scalable synthetic systems can control for variability across contexts and isolate a single biological variable to measure, but they have technical limitations. For example, across effectors that function as activators, repressors or both, we generally find stronger effects in K562 versus HEK293T cells, and it seems possible that a global effect, for example, on rTetR recruitment strength, contributes to this result. While these cell lines are useful for screening and we validated some effectors in other cells including J774 macrophages, THP-1 monocytes, primary T cells and WTC11 iPSCs, further assessment of effectors in a broader set of cell types, including primary and stem cells, will be important to uncover dependencies that are particularly relevant in disease and development. We also found that dCas9 HT-recruit screens varied in their dynamic range depending on the sgRNA and target. Measurements in additional endogenous target contexts^{102,103}, including sgRNAs with varied proximity to target TSS and enhancers, will be needed to dissect the complex relationships between target context and effector function. While 80 amino acid domain libraries are powerful due to their scalability, programmability, focus on short functional regions and the deliverability of the compact domains, using longer sequences could uncover additional forms of context dependence as longer sequences are more likely to encode multiple functions and layers of regulation. Isolating domains can reveal functions that are masked in the full-length context, but these functions can also be unrelated to the protein's normal function. Although termed transcriptional effectors, these assays do not directly measure mRNA synthesis, and it is still possible that some hits affect processes downstream of transcription, such as mRNA maturation. Toward understanding the mechanisms behind effector context dependence, it would next be useful to better measure and manipulate the endogenous factors that co-occupy the targeted promoters and enhancers.

Overall, our work highlights the utility of a high-throughput process for protein engineering in synthetic biology that is enabled by the ability to synthesize and test large pooled libraries of domains for function directly in human cells. In the future, HT-recruit could be adapted to systematically discover and characterize effectors for the other forms of gene and epigenetic editing, including base and prime editing¹⁰⁴.

Online content

Any methods, additional references, Nature Portfolio reporting summaries, source data, extended data, supplementary information, acknowledgements, peer review information; details of author contributions and competing interests; and statements of data and code availability are available at <https://doi.org/10.1038/s41587-024-02442-6>.

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Methods

Cell lines and cell culture

Experiments presented here were carried out in K562 (American Type Culture Collection (ATCC), CCL-243, female), HEK293T (ATCC, CRL-3216), LentiX-293T (Takara Bio, 632180), primary human CD8⁺ T cells, NALM6 (ATCC), MCF10a (ATCC), THP-1 (ATCC), WTC11 (Gladstone Institute) and J774 (ATCC) cells. Cells were cultured in a controlled humidifier at 37 °C and 5% CO₂. K562 cells were cultured in RPMI 1640 (Gibco, 11-875-119) media supplemented with penicillin (10,000 IU ml⁻¹), streptomycin (10,000 µg ml⁻¹), 2 mM L-glutamine and 10% Tet-approved FBS (Omega Scientific, 20014T). HEK293T or LentiX cells were maintained in Dulbecco's modified Eagle medium (DMEM; Gibco, 10569010) supplemented with 25 mM D-glucose (Gibco), 1 mM sodium pyruvate (Gibco), 1× GlutaMAX (Gibco), penicillin (10,000 IU ml⁻¹), streptomycin (10,000 µg ml⁻¹) and 10% Tet-approved FBS (Omega Scientific, 20014T). When HEK293T cells reached 80% confluence, they were gently washed with 1× Dulbecco's phosphate-buffered saline (DPBS) (Life Technologies) and passaged using 0.25% trypsin (Life Technologies). MCF10a cells were maintained in DMEM/F12 (Invitrogen, 11330-032) supplemented with insulin (10 µg ml⁻¹), cholera toxin (100 ng ml⁻¹), hydrocortisone (0.5 mg ml⁻¹), EGF (20 ng ml⁻¹), penicillin (10,000 IU ml⁻¹), streptomycin (10,000 µg ml⁻¹) and 5% horse serum (Invitrogen, 16050-122). J774 cells were cultured in 10 cm plates in DMEM media supplemented with 2 mM glutamine, 100 U ml⁻¹ penicillin, 100 µg ml⁻¹ streptomycin and 10% heat-inactivated FBS and were passaged two to three times weekly by exchanging media when cells reached ~90% confluency, incubating for 24 h and scraping. For long-term storage, cells were resuspended in freezing media (10% dimethyl sulfoxide (Sigma-Aldrich) and cell media) in a cryovial and frozen at -80 °C. THP-1 cells were cultured in RPMI 1640 (Gibco, 11-875-119) media supplemented with penicillin (10,000 IU ml⁻¹), streptomycin (10,000 µg ml⁻¹), 2 mM L-glutamine and 10% heat-inactivated FBS (Omega Scientific, 20014T). WTC11 (Gladstone Institute) human iPSCs were cultured in mTeSR Plus medium (STEMCELL Technologies) and cultured on Geltrex-coated plates (Gibco). Cells were routinely passaged at approximately 80% confluency by enzymatic lifting with Accutase (Innovative Cell Technologies), and media was supplemented with 10 µM Y27632 (MedChemExpress) for the first 24 h after passaging. Primary CD8⁺ T cells were isolated using the EasySep Human CD8⁺ T cell Isolation Kit (STEMCELL Technologies) from the whole peripheral blood provided by healthy donors at the Blood Donor Center at Boston Children's Hospital through a protocol approved by the Boston University Institutional Review Board. Cells were activated by Human T-activator CD3/CD28 for T cell expansion and activation Dynabeads (Gibco, 11131D) and cultured in X-Vivo 15 Medium (Lonza, 04-418Q) supplemented with 5% human AB serum (Valley Biomedical, HP1022), 10 mM N-acetyl L-cysteine (Sigma-Aldrich, A9165), 55 µM 2-mercaptoethanol (Gibco) and 100 U ml⁻¹ interleukin 2 (PeproTech, 200-02). NALM6 target cell lines were cultured in RPMI1640 medium supplemented with 5% FBS, 1% L-glutamine (Gibco, A2916801) and 1% penicillin-streptomycin.

Generating reporter cell lines

The minCMV, human pEF1α from the *EEF1A1* gene, RSV, human UbC and human PGK promoters were selected to span a range of basal expression levels. To include more minimal promoters with low basal expression, two promoters (NTX and NT21, which both lack a TATA box, in contrast to minCMV) were selected from a published resource¹⁰⁵. These promoters were each cloned into a reporter plasmid with homology arms for integration into the AAVS1 safe harbor locus (Supplementary Table 4).

Reporter cell lines were generated by transcription activator-like effector nuclease (TALEN)-mediated homology-directed repair via integration of a donor construct into the AAVS1 locus of cells as follows: 1.2×10^6 K562 cells were electroporated in Amasa solution

(Lonza Nucleofector 2b, setting T-16), or 8×10^4 HEK293T cells were transfected using Lipofectamine LTX (Invitrogen), according to manufacturer's instructions, with 1,000 ng of reporter donor plasmid and 500 ng of each TALEN-L (Addgene, 35431) and TALEN-R (Addgene, 35432) plasmid (targeting upstream and downstream the intended DNA cleavage site, respectively). Cells were selected with 500 ng ml⁻¹ puromycin (InvivoGen) starting 48 h after electroporation/transfection for 5 days or until all of the negative control cells died. The TALEN plasmids were gifts from F. Zhang¹⁰⁶. These cells were not authenticated.

Domain library design and cloning

The nuclear protein Pfam domain library was designed previously³². Briefly, we queried the UniProt database¹⁰⁷ for human genes that can localize to the nucleus. We then retrieved Pfam-annotated domains using the ProDy searchPfam function¹⁰⁸. We filtered for domains that were 80 amino acids or shorter and excluded the C2H2 zinc finger DBDs, which are highly abundant, repetitive and not expected to function as transcriptional effectors. We retrieved the sequence of the annotated domain and extended it equally on either side to reach 80 amino acids total. Duplicate sequences were removed, and then codon optimization was performed for human codon usage, removing BsmBI sites and constraining GC content to between 20% and 75% in every 50 nucleotide window using DNA Chisel¹⁰⁹. A total of 499 random controls of 80 amino acids lacking stop codons were computationally generated as controls. A total of 362 elements tiling the DMD protein in 80 amino acid tiles with a 10 amino acid sliding window were also included as controls because DMD was not thought to be a transcriptional regulator. In total, the library consists of 5,954 elements.

The CRTF tiling library was previously designed³⁴. Briefly, 735 chromatin regulators and 1294 TF proteins were tiled with 80 amino acid tiles with a 10 amino acid tile sliding window. In addition, 2,028 random sequence controls (lacking stop codons) were computationally generated. Duplicate sequences were removed, sequences were codon optimized for human codon usage, 7xC homopolymers were removed, BsmBI restriction sites were removed, rare codons (less than 10% frequency) were avoided and the GC content was constrained to be between 20% and 75% in every 50 nucleotide window (performed with DNA Chisel¹⁰⁹). The library includes a total of 128,565 elements (including small sublibraries that were not analyzed here), which were split across five subpools that were separately amplified and cloned into the dCas9 recruitment plasmid pJT216.

Oligonucleotides with lengths of 300 nucleotides to encode these protein sequences and flanking cloning and amplification adapters were synthesized as pooled libraries (Twist Biosciences; Supplementary Table 6). The methods used to amplify these oligonucleotides, clone them into a lentiviral dCas9 recruitment vector pJT216 (Addgene, 187320; Supplementary Table 5), produce lentiviral libraries and deliver them to cells are described in detail in Supplementary Note 4.

We also used previously cloned plasmid libraries wherein these domains were cloned onto the rTetR or rTetR(SE-G72P) mutant version of rTetR with reduced leakiness^{32,34,110}. In addition, we integrated previous HT-recruit datasets that also used these previously cloned libraries, with rTetR used for the Pfam library at the pEF reporter in K562 cells and rTetR(SE-G72P) used for all other screens.

HT-recruit to measure repressor activity at reporters

For the nuclear Pfam domain repressor screens, 4×10^7 K562 PGK or UbC reporter cells per replicate were infected with 72 ml of the lentiviral library by spinfection. Cells were started in T150 flasks. Two days later, cells were ~10% mCherry⁺ by flow cytometry (ZE5 Cell Analyzer), and selection began with 10 µg ml⁻¹ blasticidin (InvivoGen) in T225 flasks. Cells were maintained in log growth conditions each day by diluting cell concentrations back to 5×10^5 cells per ml, with at least 1×10^8 cells total remaining per replicate, such that the lowest maintenance coverage was >10,000× cells per library element. On day 7 postinfection,

recruitment was induced by treating the cells with 1,000 ng ml⁻¹ doxycycline (Tocris Bioscience) for 5 days, and then cells (>5,000× coverage) were collected for magnetic separation.

HEK293T screens were performed in T225 flasks and split daily via dissociation with 1× TrypLE (Gibco). For HEK293T cells with the pEF reporter, 1.25 × 10⁷ cells per replicate were infected by spinfection with 25 ml of virus in six-well plates with 5 ml per well, resulting in 85% mCherry⁺ 2 days later. Two days after infection, 1 µg ml⁻¹ doxycycline and 10 µg ml⁻¹ blasticidin were added. Four days later, 75% of cells (~6,500× coverage) were collected for magnetic separation, and the remaining cells were washed twice and resuspended in fresh media. Eight days later, those cells (10,000× coverage) were collected for magnetic separation.

For HEK293T cells with the UbC reporter, 1.65 × 10⁷ cells per replicate were infected by overnight incubation with varied doses of virus in T225 flasks. The screen continued with the high dose (4.13 ml), resulting in ~24% mCherry⁺ cells 2 days later. Two days after infection, 10 µg ml⁻¹ blasticidin was added. Five days later, the cells were frozen. Later, cells were thawed, and then blasticidin was added and refreshed daily for 6 days. Then, 1 µg ml⁻¹ doxycycline was added, and 4 days later, cells (>6,500× coverage) were collected for magnetic separation followed by sequencing (described in detail in Supplementary Note 4).

HT-recruit to measure activation activity at reporters

For the nuclear Pfam domain activator screens, 4 × 10⁷ K562 NTX or NT21 reporter cells per replicate were infected with 72 ml of the lentiviral library by spinfection. Cells were started in T150 flasks. Two days later, cells were ~10% mCherry⁺ by flow cytometry (ZE5 Cell Analyzer), and selection began with 10 µg ml⁻¹ blasticidin (InvivoGen) in T225 flasks. Cells were maintained in log growth conditions each day by diluting cell concentrations back to 5 × 10⁵ cells per ml, with at least 1 × 10⁸ cells total remaining per replicate, such that the lowest maintenance coverage was >10,000× cells per library element. On day 7 postinfection, recruitment was induced by treating the cells with 1,000 ng ml⁻¹ doxycycline (Tocris Bioscience) for 2 days, and then cells (>6500× coverage) were collected for magnetic separation.

The protocol was similar for HEK293T cells, but cells were maintained daily in a T225 flask at >1.4 × 10⁷ via dissociation with 1× TrypLE (Gibco). For HEK293T cells with the minCMV and silenced PGK reporters, 1.65 × 10⁷ cells per replicate were infected by overnight incubation with varied doses of virus in T225 flasks. The screen continued with the high dose (4.13 ml), resulting in ~24% mCherry⁺ cells 2 days later. Two days after infection, 10 µg ml⁻¹ blasticidin was added. The minCMV screen continued—5 days later, 1 µg ml⁻¹ doxycycline was added, and 2 days later, the cells (>2,000× coverage) were collected for magnetic separation. Meanwhile, 5 days after blasticidin addition, PGK cells were frozen. Later, PGK cells were thawed, and then blasticidin was added and refreshed daily for 6 days. Then, 1 µg ml⁻¹ doxycycline was added, and 2 days later, cells (>6,500× coverage) were collected for magnetic separation followed by sequencing (described in detail in Supplementary Note 4). Additionally, cells from the HEK293T minCMV reporter screen were collected for FLAG staining to measure fusion protein expression levels (Supplementary Note 4).

CRISPR HT-recruit to measure transcriptional effectors at endogenous genes and reporters

CRISPR HT-recruit screens were performed with dCas9 as the DBD and a sgRNA targeting either an endogenous surface marker (CD2, CD20, CD28 or CD43) or the TetO site upstream a reporter with the synthetic surface marker (TetO-minCMV or TetO-pEF1α; Supplementary Table 9). First, the sgRNA was stably delivered to K562 cells by lentivirus and selected with puromycin for 3–4 days. The cells were confirmed to be >95% mCherry⁺ by flow cytometry (BD Accuri C6).

For the first replicate of the CD2 (sg717 and sg718) and CD43 screens, 1.35 × 10⁷ of these cell lines per replicate were infected with

20 ml of the 4×-concentrated (LentiX) lentiviral dCas9-Pfam library by spinfection (split and spun in 2 × 10 ml volumes in 50 ml Falcon tubes). The cells were ~10% BFP⁺ by flow cytometry (ZE5) after 5 days (as measured from a set-aside plate that did not receive antibiotic selection). For the CD20, CD28, other CD2 sgRNAs and the second replicate of the CD2 sg717 screens, 1.35 × 10⁷ of these cell lines per replicate were infected with 20 ml of the 4×-concentrated (LentiX) lentiviral dCas9-Pfam library by spinfection in 4 ml per well in six-well plates. The cells were ~68% BFP⁺ by flow cytometry (ZE5) after 5 days. For the TetO sgRNA and the second replicate of the CD43 screens, 1.5 × 10⁷ cells were spininfected with 9 ml of 4×-concentrated virus split in three wells of a six-well plate, resulting in ~50% BFP⁺ cells.

For all of these samples, cells were maintained in T175 flasks. Two to three days after infection, selection began with 10 µg ml⁻¹ blasticidin (InvivoGen). Cells were maintained in log growth conditions each day by diluting cell concentrations back to 5 × 10⁵ cells per ml (and replenishing blasticidin), with at least 4.2 × 10⁷ cells total remaining per replicate, such that the lowest maintenance coverage was >5,000× cells per library element. On day 10 postinfection, cells (>20,000× coverage) were collected for magnetic separation.

For the dCas9-CRTF screens, lentivirus for the library was generated using 16 × 15 cm dishes of HEK293T cells and then concentrated 4× using LentiX. Then 1.15 × 10⁸ K562-sgRNA cells (sg717 or sg15) per replicate were infected with 72 ml of the lentiviral library by spinfection for 2 h with two separate biological replicates of the infection, resulting in 18–23% BFP⁺ cells in unselected cells after 4 days. Two days after infection, the cells were selected with 10 µg ml⁻¹ blasticidin (InvivoGen). Cells were >95% BFP⁺ by the final time point. On day 11 postinfection, 5 × 10⁸ cells (>3,000× coverage) were collected for magnetic separation.

An additional growth-based CRISPR HT-recruit screen was performed with the Pfam domain library and a sgRNA targeting the eGATA1 enhancer (which has a fitness effect when repressed) or a safe-targeting negative control with no fitness effect. For the first replicate, 1.35 × 10⁷ cells of these sgRNA cell lines per replicate were infected with 20 ml of the 4×-concentrated (LentiX) lentiviral dCas9-Pfam library by spinfection (split and spun in 2 × 10 ml volumes in 50 ml Falcon tubes). Cells were maintained in T175 flasks. The cells were ~10% BFP⁺ by flow cytometry (ZE5) after 5 days (as measured from a set-aside plate that did not receive antibiotic selection). Three days after infection, cells were selected with 10 µg ml⁻¹ blasticidin (InvivoGen) for 7 days. The second replicate was performed later, with 1.5 × 10⁷ cells spininfected with 9 ml of virus split in three wells of a six-well plate, resulting in ~60% BFP⁺ cells. Cells were maintained in log growth conditions each day by diluting cell concentrations back to 5 × 10⁵ cells per ml (and replenishing blasticidin), with at least 4.2 × 10⁷ cells total remaining per replicate, such that the lowest maintenance coverage was >5,000× cells per library element. On day 13 postinfection, cells (>20,000× coverage) were collected for genomic DNA extraction and sequencing (described in Supplementary Note 4). Additionally, screen cells with safe N4293 and CD43 sg15 were collected for FLAG staining to measure fusion protein expression levels (Supplementary Note 4).

HT-recruit analysis

The HT-recruit-Analyze pipeline¹¹¹ was used to run Bowtie to align Illumina sequencing reads to an index of domain-encoding sequences and compute effector enrichment scores. Sequencing reads were trimmed to remove the 19–24 bp stagger plus constant primer handle region from the beginning of the read. For most samples, reads were further trimmed to retain only the first 131 bp of the domain-encoding sequence. Alignment parameters were optimized for a high alignment rate (>80%) and a low ambiguous alignment rate (<1%). With these metrics, for the Pfam library, mismatch tolerance was set to Bowtie -m 3 (as each domain is codon optimized so they can differ at more DNA base pairs than amino acid residues), while for the tiling library,

no mismatches were allowed. This mismatch tolerance approach is tailored toward ignoring sequencing errors while removing reads with synthesis or PCR errors because it allows mismatches if the sum of their *Q* scores is <30 and the number of mismatches in the initial 28 bp is less than -*m*.

Then, the $\log_2(\text{OFF:ON})$ score was computed using sample depth-normalized counts in the OFF (unbound) and ON (bound) samples. To avoid inaccurate measurements from low coverage, if an element had fewer than five counts in one sample (bound or unbound fraction), its count was set to five. If an element had fallen below that threshold in both samples, it was removed from the data tables. For downstream analyses, the Pfam and tiling libraries were further filtered for elements with >5 or >20 counts, respectively, in both samples.

Analysis was performed similarly for the FLAG-based expression level measurements, using the FLAG high and low samples. An element was defined as well-expressed if its $\log_2(\text{FLAG high:low})$ score was greater than 1 s.d. above the median of the random controls.

Then, an element from the Pfam library was defined as an activator or repressor hit if its $\log_2(\text{OFF:ON})$ score was 2 s.d. below (for activators) or above (for repressors) the median of the poorly expressed domains (as defined by FLAG measurements with that DBD and cell type). This approach provided a conservative hit threshold that was chosen to identify robust effectors, but subthreshold domains may also have some effector activity. For some analyses of the pEF1 α rTetR screen in K562 cells, we also report a lower threshold (0.9), equivalent to the strength of the weakest repressor (DUF1087) that was individually validated³². The tally of domains that are hits in both replicates of a screen excludes growth-based screens, screen memory time points after doxycycline washout and screens with one replicate (sgCD2_718, sgCD20_135, sgCD20_148, sgCD20_275, sgCD28_07, sgCD28_56, sgCD28_94, sgCD2_39, sgCD2_42 and sgCD2_89). Additional analysis of the tiling library is described in Supplementary Note 4.

CRISPRi effector benchmarking growth screens

Detailed descriptions of the methods for three CRISPRi benchmarking screens using growth readouts are in Supplementary Note 4.

Individual recruitment assays

The methods used to perform individual recruitment assays are described in detail in Supplementary Note 4. In total, these were performed (1) by transient transfection or lentiviral delivery; (2) with rTetR, dCas9, dCas12a, dCasMINI, MS2, rapamycin-inducible ZFHD1 or synZiFTR recruitment; (3) in K562, HEK293T, THP-1, J774, iPSC or T cells.

Western blot for effector fusion expression

K562 cells were transduced with a lentiviral vector containing a dCas9-3XFLAG-effector-T2A-BFP-BSD and 3 days later selected with blasticidin (10 $\mu\text{g ml}^{-1}$) for 15 days until complete. Greater than 80% of the cells appeared BFP⁺ by microscopy. In total, 5–10 million cells were lysed in lysis buffer (1% Triton X-100, 150 mM NaCl, 50 mM Tris (pH 7.5) and protease inhibitor cocktail). Protein amounts were quantified using the Pierce BCA Protein Assay Kit (Bio-Rad). Equal amounts were loaded onto a gel and transferred to a PVDF membrane. The membrane was probed using FLAG M2 monoclonal antibody (mouse; Sigma-Aldrich, F1804; 1:1,000) and β -actin antibody (rabbit; Abcam, ab8227; 1:1,000) as primary antibodies. Antirabbit 680 (IRDye 680LT donkey antirabbit IgG secondary antibody; Li-Cor, 926-68023) and antimouse 800 (IRDye 800CW goat anti-mouse IgG secondary antibody; Li-Cor, 926-32210) at 1:10,000 dilution were used as secondary antibodies. Blots were imaged on a Li-Cor Odyssey CLx. Band intensities were quantified using ImageJ¹¹².

Chromatin modification mapping and RNA-seq

Stable K562 cell lines expressing dCas9 fusions were analyzed by CUT&RUN and RNA-seq (details are given in Supplementary Note 4).

Analysis software for flow cytometry data

The flow cytometry data were analyzed with the MATLAB program EasyFlow (<https://antebilab.github.io/easyflow/>) or the Python program Cytoflow (<https://cytoflow.github.io/>).

Statistical analyses

Nonlinear regression, the Kruskal–Wallis test and two-way analysis of variance were performed in Prism 9.3.1. Other statistical analyses were performed in Python using SciPy¹¹³. No methods were used to determine whether the data met the assumptions of the statistical approach.

Reporting summary

Further information on research design is available in the Nature Portfolio Reporting Summary linked to this article.

Data availability

All Illumina sequencing data generated in this study are deposited in the National Center for Biotechnology Information Sequence Read Archive database (PRJNA1160796)¹¹⁴. Processed data are given in Supplementary Tables 1–3 (<https://doi.org/10.5281/zenodo.13756269>)¹¹⁵. Plasmids generated in this study are available from Addgene (<https://www.addgene.org/browse/article/28228872/>). An uncropped scan of the western blot is provided as source data. Source data are provided with this paper.

Code availability

Code to analyze HT-recruit domain screens (<https://github.com/bintulab/HT-recruit-Analyze>) and sgRNA screens (<https://github.com/elifesciences-publications/dmorgens-castle>) is available online.

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Author contributions

J.T., M.C.B. and L.B. designed the overall study. J.T., M.V.V., A. and C.L. generated reporter cell lines. J.T. and K.S. cloned pooled libraries.

J.T., M.V.V. and A. performed HT-recruit, other screens and individual validations. G.H. designed and tested sgRNAs. A. and K.L. designed and performed dCas12a experiments. J.T., D.Y., N.D., A.X.M. and P.H.S. designed oligonucleotide libraries. P.H.S. characterized homeodomain mutants. J.T., A. and D.Y. performed CRISPRi benchmarking screens. M.G. and R.A.K. performed experiments with J774 cells. K.L. performed experiments with MCF10a cells. R.V. and C.Z. performed experiments with THP-1 cells. A.V.-J. and S.A. performed experiments with iPSC. A. and R.V. performed RNA-seq. X.X. designed and performed dCasMINI experiments with support from L.S.Q. J.T., V.K. and P.P.D. designed and performed MS2-recruitment experiments. H.Y. designed, performed and analyzed synZiFTR T cell experiments with support from A.S.K. and assistance from T.S.O. J.T. and M.V.V. analyzed the data and prepared the figures, with additional analysis of the tiling library by N.D. J.T., M.V.V., L.B. and M.C.B. wrote the manuscript with input from all authors.

Competing interests

J.T., L.B. and M.C.B. are inventors on provisional patents related to this work and acknowledge outside interest in Stylus Medicine. L.S.Q. is a founder and scientific advisory board member of Epicrispr Biotechnologies and Refuge Biotechnologies. X.X. and L.S.Q. are inventors on provisional patents related to dCasMINI. A.S.K. is an inventor on patents related to synZiFTR, is a scientific advisor for and holds equity in Senti Biosciences and Chroma Medicine and is a cofounder of K2 Biotechnologies. All other authors declare no competing interests.

Additional information

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